



Electrical stimulation-based bone fracture treatment, if it works so well why do not more surgeons use it?

Mit Balvantray Bhavsar¹ · Zhihua Han¹ · Thomas DeCoster² · Liudmila Leppik¹ · Karla Mychellyne Costa Oliveira¹ · John H Barker¹

Received: 24 November 2018 / Accepted: 29 March 2019 / Published online: 6 April 2019
© Springer-Verlag GmbH Germany, part of Springer Nature 2019

Abstract

Background Electrical stimulation (EStim) has been proven to promote bone healing in experimental settings and has been used clinically for many years and yet it has not become a mainstream clinical treatment.

Methods To better understand this discrepancy we reviewed 72 animal and 69 clinical studies published between 1978 and 2017, and separately asked 161 orthopedic surgeons worldwide about their awareness, experience, and acceptance of EStim for treating fracture patients.

Results Of the 72 animal studies, 77% reported positive outcomes, and the most common model, bone, fracture type, and method of administering EStim were dog, tibia, large bone defects, and DC, respectively. Of the 69 clinical studies, 73% reported positive outcomes, and the most common bone treated, fracture type, and method of administration were tibia, delayed/non-unions, and PEMF, respectively. Of the 161 survey respondents, most (73%) were aware of the positive outcomes reported in the literature, yet only 32% used EStim in their patients. The most common fracture they treated was delayed/non-unions, and the greatest problems with EStim were high costs and inconsistent results.

Conclusion Despite their awareness of EStim's pro-fracture healing effects few orthopedic surgeons use it in their patients. Our review of the literature and survey indicate that this is due to confusion in the literature due to the great variation in methods reported, and the inconsistent results associated with this treatment approach. In spite of this surgeons seem to be open to using this treatment if advancements in the technology were able to provide an easy to use, cost-effective method to deliver EStim in their fracture patients.

Keywords Bone fracture healing · Electrical stimulation treatment · Literature review · Survey of orthopedic surgeons

Introduction

The earliest report of using EStim to treat bone fractures in patients appeared in the mid-1800s in which Garrat [1] described using metallic needles placed in non-healing fractures to deliver DC EStim, that resulted in successful

healing. Today, in the clinical setting EStim is administered using three different approaches; direct current (DC), pulsed electromagnetic field (PEMF), and capacitive coupled (CC). DC EStim is administered via a surgically implanted EStim power source and electrodes, and is administered at dosages between 10 and 100 μ A of current [2]. CC and pulsed PEMF are both administered externally. In CC an alternating voltage is applied to cutaneous electrodes placed on opposite sides of the fracture generating an electrical field of 0.1–20 G [3]. In PEMF alternating currents, in current-carrying coils, on the skin over the fracture site, generate a pulsed electromagnetic field ranging between 3 and 10 V peak-to-peak within the fracture site [4].

In most cases EStim is used as a last resort after other treatments have failed and/or in combination with other treatments in cases of problematic fractures that heal slowly (delayed union) or do not heal at all (non-union) [5].

Mit Balvantray Bhavsar and Zhihua Han contributed equally to the work.

✉ John H Barker
JHB121654@gmail.com

¹ Frankfurt Initiative for Regenerative Medicine, Experimental Trauma and Orthopedic Surgery, J.W. Goethe-University, Friedrichsheim gGmbH, Haus 97 B, 1OG, Marienburgstraße. 2, 60528 Frankfurt am Main, Germany

² Department of Orthopedics and Rehabilitation, University of New Mexico, Albuquerque, NM, USA

Examples include; spinal fusion [6], avascular necrosis [7], internal and external fixation [8], delayed- or non-union fractures [9], osteotomies [10], bone grafts [11], and femoral osteonecrosis [12]. In these cases, EStim has been generally reported to promote bone healing and help resolve these difficult, often chronic, costly, and debilitating fractures.

Several recently published *in vitro* studies suggest that EStim's pro-healing effect is due to its influence on the behavior and/or function of bone-forming stem cells. Along these lines, we and others have shown that EStim causes bone forming stem cells to migrate [13, 14], proliferate [15, 16], differentiate [17–20], increase mineralization [21], deposit extracellular matrix [22], attach to scaffold materials [23], and increase the expression of several osteogenic genes [19, 20]. Importantly, all these cell behaviors/functions play key roles in fracture healing and/or bone regeneration. In addition to these *in vitro* findings at the cellular level, in *in vivo* studies in rat forelimb amputation [24] and large bone defect models [25] we have demonstrated that EStim significantly stimulates new bone, cartilage, and vessel formation and promotes healing and regeneration. In spite of these positive results in preclinical and clinical studies EStim has not become a widespread, universally used clinical treatment.

To better understand this discrepancy between the reported positive results and the relatively low use of EStim in fracture treatment we reviewed the literature and we asked orthopedic surgeons worldwide (in a survey) about their awareness, experience, and acceptance of EStim treatment in their fracture patients. Using this combined approach, we hoped to better understand the discrepancy between the demonstrated success of EStim fracture treatment, and its relatively low use clinically.

Methods

Literature review

To identify articles describing the use of EStim in bone healing, both in animal and clinical studies, we searched MEDLINE, Google Scholar, and Web of Science databases for articles describing *in vitro* and *in vivo* animal studies and clinical studies published between 1977 and 2017. To maximize the sensitivity of the search and identify the greatest number of studies, we used different combinations of the keywords “electrical stimulation” and “bone healing” and reviewed the reference lists of retrieved publications to identify additional articles we may have missed searching the three databases. We categorized the total number of articles identified into “animal studies”, “clinical studies”, “cell/organ *in vitro*”, and “reviews/meta analyses” (Table 1). Since the focus of our study was to investigate EStim's effect on fracture healing we reviewed only articles that described fracture healing in animal and clinical

Table 1 Total publications identified using different combinations of the keywords “electrical stimulation” and “bone healing”

| Study type | Number of publications |
|----------------------------|------------------------|
| Animal studies | 72 |
| Clinical studies | 69 |
| Cell/organ <i>in vitro</i> | 238 |
| Review/meta-analysis | 53 |
| Total | 432 |

studies, and excluded publications focused on *in vitro* studies, electrical properties of bone, connective tissue, electrical stimulation of nerves and reviews or meta-analyses. The animal studies we identified and reviewed are listed in Table 2 categorized by animal model studied, bone and fracture type, type of EStim treatment used, outcomes, and the listing of the published article, along with the number of occurrences in each category. The clinical studies reviewed are listed in Table 3 under the subtitles; bone and fracture type, number of cases, EStim treatment used, outcomes, complications, and the published article cited along with the total numbers for each of these categories. The total number for each of these categories is summarized in Table 3. The language of the publications reviewed was English.

Orthopedic surgeon survey

To determine the level of awareness, experience, and acceptance of EStim-based bone fracture treatment we asked orthopedic surgeons six questions (listed in Figs. 1, 2, 3, 4, 5, 6) using a closed online automated survey method (Survey-Monkey software, Palo Alto, USA). Survey participants were identified from our own network of colleagues and based on their surgical specialty, “Orthopedic Surgeons” in the online professional networking website, LinkedIn [26]. Between May and August 2017, a total of 620 invitations were emailed to orthopedic surgeons worldwide, and their IP addresses were used to record their country of origin and to prevent duplicate entries. No other personal information was collected or stored from the respondents. With this online survey method participants were allowed to review their responses prior to submitting their completed survey. Incomplete surveys were not included in this analysis.

Results

Literature review

Our initial literature searches identified a total of 432 articles, published between 1977 and 2017 that focused on the use of EStim to promote bone growth, fracture healing, and

Table 2 Publications between 1977 and 2017 describing animal studies that use EStim to treat bone fractures

| Animal model | Bone affected and/or fracture type | Type of EStim treatment | Outcome | Published article |
|--------------|------------------------------------|--|------------------------|------------------------|
| Rabbits | Femur/osteotomy | Type: PEMF Settings: 220–260 G | Improved healing | Aydin and Bezer [46] |
| | Tibia/osteotomy | Type: PEMF | Improved healing (69%) | Barak et al. [47] |
| | Tibia/fracture | Type: PEMF Settings: pulse width 85 μ s Duration: 30 min/day | No effect | Buzza et al. [48] |
| | Tibia/osteotomy | Type: PEMF Settings: time-varying field 1.5 Hz Duration: 1 h/day | Improved healing | Fredericks et al. [49] |
| | Lumbar spine/fusion | Type: DC Settings: 20–60 μ A | Improved healing | France et al. [50] |
| | Lumbar spine/fusion | Type: CC | Improved healing | Gilotra et al. [51] |
| | Patella–tendon junction/fracture | Type: CC Settings: 1.5–2.5 mA | Improved healing | Hu et al. [52] |
| | Mandible/defect | Type: DC Settings: 20 μ A Duration: 4 weeks continuous | Improved healing | Kim et al. [53] |
| | Femur/defect | Type: PEMF Settings: 0.8 mT Duration: 4 h/day | Improved healing | Matsumoto et al. [54] |
| | Tibia/fracture | Type: PEMF Settings: 8 mT; 50 Hz Duration: 0.5 h/day | Improved healing | Ottani et al. [55] |
| | Tibia/fracture | Type: DC Settings: 20 μ A Duration: 0.5 h/day continuous | Improved healing | Rubinacci et al. [56] |
| | Mandible/defect | Type: DC Settings: 7 μ A Duration: 1–2 weeks continuous | No effect | Shafer et al. [57] |
| | Tibia diaphysis/fracture | Type: PEMF Settings: 1.8 G; 1.5 Hz | Improved healing | Shimizu et al. [58] |
| | Femur, tibia/fracture | Type: PEMF Settings: repetitive pulse-72 Hz Duration: 12 h/day continuous | No effect | Smith and Nägele [59] |
| | Tibia/osteotomy | Type: PEMF Settings: asymmetric pulse 1.5 Hz Duration: 20 days continuous | No effect | Taylor et al. [60] |
| | Knee/osteochondral lesion | Type: PEMF Settings: 1.5 mT; 75 Hz Duration: 4 h/day for 40 days | Improved healing | Veronesi et al. [61] |
| | Humerus/fracture | Type: PEMF Settings: 2 G, 25 μ s pulses at 10 Hz Duration: 12 h/day \times 14 days | Improved healing | Yonemori et al. [62] |
| | Tibia/fracture | Type: DC Settings: 1, 5, 20 μ A | Improved healing | Zimmermann et al. [63] |

Table 2 (continued)

| Animal model | Bone affected and/or fracture type | Type of ESstim treatment | Outcome | Published article |
|--------------|------------------------------------|---|---------------------------|-------------------------|
| Dogs | Ulna/fracture | Type: DC Settings: 20 μ A Duration: continuous | No effect | Berry et al. [64] |
| | Tibia/fracture | Type: DC Settings: 10–20 μ A Duration: continuous | Improved healing (70–80%) | Bins-Ely et al. [65] |
| | Mandible/defect | Type: DC Settings: 20 μ A | No effect | Branham et al. [66] |
| | Radius/fracture | Type: DC Settings: 0.1–17 μ A | Improved healing | Chakkalakal et al. [67] |
| | Femur/fracture | Type: DC Settings: 50 μ A Duration: 6 weeks continuous | Improved healing | Colella et al. [68] |
| | Radius, ulna/fracture | Type: DC Settings: 20 μ A Duration: 12 weeks continuous | Improved healing | Connolly et al. [69] |
| | Lumbar spine/fusion | Type: DC Settings: 0.83–10 μ A Duration: 6 weeks continuous | Improved healing | Dejardin et al. [70] |
| | Femur/fracture | Type: CC Settings: biphasic waveforms | Improved healing | Doyle [71] |
| | Radius/fracture | Type: DC Settings: 3–5 μ A | Improved healing | Fuentes et al. [72] |
| | Tibia/fracture | Type: PEMF Settings: 0–2.4 G Duration: 4 h/day | Improved healing | Inoue et al. [73] |
| | Periodontal/defect | Type: DC Settings: 3–6 nA Duration: continuous | No effect | Jacobs and Norton [74] |
| | Ulna/non-union | Type: DC Settings: 20 μ A | Improved healing (22%) | Jacobs et al. [75] |
| | Lumbar spine/fusion | Type: PEMF Settings: 1 G; 1.5 Hz Duration: 0.5–1 h/day | No effect | Kahanovitz et al. [76] |
| | Femur/fracture | Type: DC Settings: 20 μ A | No effect | Lindsey et al. [77] |
| | Cranium/osteogenesis | Type: DC Settings: 20 μ A | No effect | Moderessi et al. [78] |
| | Mandible/osteogenesis | Type: PEMF Duration: 1 h/day | Improved healing | Ortman et al. [79] |
| | Mandible/non-union | Type: DC | Improved healing | Park et al. [80] |
| | Tibia/non-union | Type: DC Settings: 20 μ A | Improved healing | Paterson et al. [81] |
| | Tibia/non-union | Type: DC Settings: 20 μ A | Improved healing | Paterson et al. [82] |
| | Tibia/defect | Type: DC Settings: 0.2–20 μ A | Improved healing | Paterson et al. [83] |

Table 2 (continued)

| Animal model | Bone affected and/or fracture type | Type of EStim treatment | Outcome | Published article |
|--------------|------------------------------------|---|------------------|------------------------------|
| | Tibia/fracture | Type: CC Settings: 3–6.3 V; 60 kHz Duration: continuous—28 days | No effect | Pepper et al. [84] |
| | Hip prostheses | Type: CC Settings: 5–6 V; 60 kHz | No effect | Schutzer et al. [85] |
| | Mandible/defect | Type: DC Settings: 20 μ A | Improved healing | Shayesteh et al. [86] |
| | Femur/osteotomy | Type: DC Settings: 1.5 V | Improved healing | Shokry [87] |
| | Tibia, femur/fracture | Type: DC Settings: 0–50 μ A | Improved healing | Srivastava [88] |
| | Femur/fracture | Type: PEMF Settings: 1.5 mT Duration: 6 h/day | Improved healing | Atalay et al. [89] |
| Rats | Tibia/osteoporosis | Type: CC Settings: low voltage; 60 Hz | Improved healing | Brighton et al. [90] |
| | Tibia/fracture | Type: CC Duration: 20 min/day | Improved healing | Giannunzio et al. [91] |
| | Spine/fusion | Type: PEMF Duration: 18 h/day | Improved healing | Guizzardi et al. [92] |
| | Tibia/osteoporosis | Type: PEMF Settings: 1 G; 5 ms pulse; 15 Hz Duration: 2 h/day | No effect | Jagt et al. [93] |
| | Tibia/osteoporosis | Type: PEMF Settings: 30 mW/cm ² ; 1.5 MHz | Improved healing | Lirani-Galvao et al. [94] |
| | Tibia/osteoporosis | Type: CC Settings: 10 V peak-peak Duration: 2 h/day | Improved healing | Manjhi et al. [95] |
| | Fibula/osteotomy | Type: CC Settings: 1590 V; 60 Hz | Improved healing | Marino et al. [96] |
| | Spine/injury | Type: CC Settings: 30 mW/cm ² | Improved healing | Medalha et al. [97] |
| | Tibia/fracture | Type: DC Settings: 20 μ A Duration: 20 min/day | Improved healing | Nakajima et al. [98] |
| | Femur/fracture | Type: PEMF Settings: 41 Gauss | Improved healing | Puricelli et al. [99] |
| | Tibia/osteoporosis | Type: PEMF Settings: 8 G; 15 Hz Duration: 2 h/day | Improved healing | Shen and Zhao [100] |
| | Spine/bone growth | Type: DC Settings: 0–100 μ A | Improved healing | Spadaro [101] |
| | Mandible/defect | Type: PEMF Settings: 1.5–1.8 G; 100 Hz | Improved healing | Takano-Yamamoto et al. [102] |
| | Periodontal/defect | Type: DC Settings: 0–100 μ A; 9 kHz Duration: once per day | Improved healing | Tomofuji et al. [103] |

Table 2 (continued)

| Animal model | Bone affected and/or fracture type | Type of EStim treatment | Outcome | Published article | |
|-----------------------|------------------------------------|---|---|----------------------------|-------------------|
| Sheep | Mandible/fracture | Type: DC Settings: 9 V Duration: 24 h | Improved healing | Uysal et al. [104] | |
| | Cranium/defect | Type: DC Settings: 2 mA; 2 Hz Duration: 15 min; 3 × weeks | Improved healing | Yang et al. [105] | |
| | Spine/injury | Type: DC (subcutaneous) Settings: 1.5 mA; 2 Hz Duration: 0.5 h/day; 3 weeks | Improved healing | Yu et al. [106] | |
| | Spine/injury | Type: DC (subcutaneous) Settings: 20–150 mA; 50 Hz Duration: 20 min/day | Improved healing | Zamarioli et al. [107] | |
| | Femur/defect | Type: PEMF Settings: 1.5 mT; 75 Hz Duration: 6 h/day | Improved healing | Benazzo et al. [108] | |
| | Tibia/fracture | Type: DC Settings: 7.5 μA Duration: 12 h/day | No effect | Dergin et al. [109] | |
| | Spine/injury | Type: DC Settings: low voltage | Improved healing | Flouty et al. [110] | |
| | Mandible/defect | Type: DC Settings: 10 μA Duration: 1 mm/day × 10 days | Improved healing | El-Hakim et al. [111] | |
| | Tibia/osteotomy | Type: PEMF Settings: 1.6 mT Duration: 24 h/day | No effect | Law et al. [112] | |
| | Tibia/fracture | Type: CC Settings: 1.5 mA; 60 kHz | Improved healing | Muthimi et al. [113] | |
| Horse | Lumbar spine/fusion | Type: DC Settings: 40–100 μA | Improved healing | Toth [114] | |
| | Metacarpus/defect | Type: PEMF Settings: 28 G; 75 Hz | Improved healing | Cane et al. [115] | |
| | Tibia/bone graft | Type: PEMF Settings: asymmetric pulse burst of 30 ms duration repeated at 1.5 Hz | Improved healing | Kold and Hickman [116] | |
| | Metatarsal-foot/osteotomy | Type: PEMF Settings: 20 G; 15 Hz Duration: 8 h/day | No effect | Sanders-Shami et al. [117] | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| Total no. of articles | Animal model (no.) | Type of bone (no.) | Type of fracture (no.) | Type of EStim (%) | Outcomes (%) |
| | Dog Rat Rabbit Sheep Horse | Tibia Femur Spine Mandible Other | Delayed-/non-union Fusion Osteotomy Large bone defects Others | PEMF DC CC | Positive Negative |
| 72 | 25 19 18 7 3 | 26 13 11 9 16 4 | 9 3 38 18 | 35 49 16 | 77 23 |

bone regeneration. Of these 432 publications, 72 reported animal studies, 69 clinical studies, 238 organ or in vitro cell culture studies, and 53 were reviews or meta-analyses (Table 1). A total of 141 publications (animal + clinical studies) were selected and reviewed, the results of which are presented herein.

Animal studies

A total of 72 animal study articles, that used EStim to treat bone fractures were reviewed. The most commonly used animal model was the dog (25), followed by rabbits (18), rats (19), sheep (7) and horses (3). In these the “bone”, and “fracture type” studied varied greatly. The bones were primarily the tibia (26), femur (13), spine (11), mandible, (9) and others (16). Some of the papers reviewed studied more than one bone. The types of fractures/pathologies were large bone defects (38), delayed- and non-unions (4), fusions (9), osteotomies (3), and others (18). The most common method used to administer electrical stimulation in the animal studies was DC EStim (49%), followed by PEMF (35%) and CC and other types, together making up 16% of the reviewed studies.

Clinical studies

A total of 69 articles describing clinical studies were reviewed, in which EStim was used to promote bone healing. The main bones treated with EStim in the clinical studies were tibia (25), femur (15), spine (15), radius (11), humerus (7) and others (20). As in the case with the animal study articles some of the clinical papers reported on more than one bone. The most common types of fractures/pathologies were delayed- and non-unions (21), spine fusions (16), arthrodeses (5), osteotomies (4), necrosis (2), large bone defects (2) and others (19). Most of the clinical studies reviewed administered EStim using PEMF (60%), followed by DC (29%), and CC and other methods (11%). The intensity of the magnetic field used in PEMF treatment ranged between 0.3 and 6 mT, while for DC the dosage was 5–40 μ A, and for CC treatment the intensity ranged between 3 and 10 V. Half (50%) of the regimens used in the clinical studies consisted of daily stimulation treatments ranging from 0.5 to 16 h/day. Nineteen of the 69 studies (27%) reported complications that included skin irritation and infections, pain [27], dislocation of the device [28], failure of the device [29] and poor patient compliance [30]. Fifty (72%) of the studies reviewed reported no complications. Finally, of the 69 clinical study publications 51 (73%) reported positive and 18 (27%) reported negative outcomes (Table 3).

Orthopedic surgeon survey

The individual questions and the responses are displayed in six separate graphs (Figs. 1, 2, 3, 4, 5, 6). Of the 620 orthopedic surgeons who were sent emails inviting them to participate in the survey, 161 (26%) from 34 countries responded. Of the 161 respondents, 44% answered that they perform more than 100, (23%) perform 51–100, (22%) perform 11–50, and (11%) perform 0–10 bone surgeries per year (Fig. 1). When asked if they were aware of published clinical studies reporting successful EStim-based fracture treatments, 85 (73%) responded “Yes” and the rest (27%) answered “No” (Fig. 2). Of the 85 respondents who said they were aware of EStim-based fracture treatments, 27 (32%) answered that they had used EStim in their fracture patients while 58 (68%) had not (Fig. 3). Of the 27 surgeons that had used EStim in their patients the pathologies they treated were mainly delayed or non-unions (61%) and large bone defects (16%). The rest, (23%) were spinal fusion, avascular necrosis, calcaneal apophysitis, Charcot foot and ankle reconstructions, loosened hip, knee prosthesis, or other types of fractures (Fig. 4). When asked what they considered to be the major problems associated with using EStim in their fracture patients, 30 (35%) identified “high cost”, 24 (28%) answered “inconsistent results”, while 8% and 5% responded that EStim devices were “impractical”, and “difficult” to use, respectively. Eleven (13%) surgeons responded that they had experienced “other” problems, and nine (11%) replied they had not experienced problems using EStim-based treatments (Fig. 5). Finally, we asked, “If an easy-to-use EStim device to treat bone fractures were available would you use it in your patients?” and 85% answered, “Yes” and the rest answered “No” (Fig. 6).

Discussion

In his review of more than 100 studies using EStim treatment, published more than 40 years ago Spadaro concludes, “About 95% are positive reports...” and goes on to qualify this assertion saying “...despite an extraordinarily wide selection of experimental techniques and models” [31]. In the present literature review of 141 papers (72 animal and 69 clinical studies), published in the 40 years since then, we also found positive results, and like Spadaro also found a great variation in bone and fracture types, treatment methods, dosages, regimens, etc., reported in the literature. The latter made it difficult to draw well-founded conclusions upon which to develop specific EStim treatment recommendations. One of the primary contributors to this confusion in the literature is the different types, dosages, and regimens used to administer EStim. In the clinical studies we reviewed

Table 3 Publications between 1977 and 2017 describing clinical studies that use EStim to treat bone fractures

| Bone affected and/or fracture type | No. of cases | Type of EStim treatment | Outcome | Complications | Published article |
|--|--------------|--|--|---|--------------------------------|
| Mandible/fracture | 12 | Type: PEMF Duration: 2 h/day × 12 days Settings: pulse duration 200 ns, rise time 8 ns, electromagnetic segment at 50 MHz and down to Hz range | No effect | Infection | Abdelrahim et al. [38] |
| Tibia/non-union | 16 | Type: CC Duration: 7–8 h/day until healed or 30 weeks Settings: 6 V peak-to-peak symmetrical sine wave signal at 63 kHz frequency | Improved healing (68%) | None reported | Abeed et al. [118] |
| Tibia/acute fracture | 106 | Type: PEMF Duration: 10 h/day × 12 weeks | No effect | None reported | Adie et al. [119] |
| Lumbar spine/fusion | 107 | Type: DC | Improved healing | None reported | Andersen et al. [6] |
| Lumbar spine/fusion | 107 | Type: DC | No effect | None reported | Andersen et al. [120] |
| Lumbar spine/fusion | 98 | Type: DC Settings: 40 and 100 µA | No effect | None reported | Andersen et al. [121] |
| Tibia/delayed- and non-union | 44 | Type: PEMF Duration: 3 h/day, maximum 36 weeks | Improved healing (77%) | None reported | Assiotis et al. [122] |
| Tibia/non-union | 9 | Type: PEMF Duration: 12–16 h/day, min 1 h/day × 48 weeks Settings: 1–5 mT peak, 5 ms burst waveform repeated at 15 Hz | No effect | One patient left the study prior to end | Barker et al. [32] |
| Femur/arthrodesis failure | 71 | Type: PEMF | Improved healing (85%) | None reported | Bassett et al. [123] |
| Tibia/fracture | 22 | Type: CC Duration: 15 h/day until healed Settings: sinusoidal wave 3–6 V at 60 kHz and 5–10 mA | No effect | None reported | Beck et al. [124] |
| Metatarsal foot/fracture | 25 | Type: CC Duration: 52 days Setting: amplitude of 3.0–6.3 V and 60 kHz frequency Effective current—5–10 mA | Improved healing (88%) | None reported | Benazzo et al. [125] |
| Femur/intertrochanteric osteotomy | 31 | Type: PEMF Duration: 8 h/day × 3 m Settings: 75 Hz, 1.3 ms impulse width, 2.5 mV amplitude, and 18 Gs magnetic field amplitude | Improved healing | None reported | Borsalino et al. [10] |
| Radius/delayed-and non-union and osteotomy | 21 | Type: PEMF Duration: 10 h/day | Improved (57% non-union healed) Improved (89% osteotomies healed) | None reported | Boyette and Herrera-Soto [126] |

Table 3 (continued)

| Bone affected and/or fracture type | No. of cases | Type of ESstim treatment | Outcome | Complications | Published article |
|---|--------------|---|---|---|----------------------------|
| Tibia/non-union | 57 | Type: DC Settings: 10–20 μ A | Improved healing (70% healed) | Eight patients did not receive adequate electricity—due to device failure | Brighton [29] |
| Tibia/non-union | 20 | Type: CC Settings: 60 kHz, 5 V peak-to-peak | No effect | 11 patients started with DC withdrew prior to end of study | Brighton and Pollack [33] |
| Tibia/non-union | 271 | Type: DC and CC | No effect | Risk factors | Brighton et al. [9] |
| Sesamoid-foot/delayed union | 1 | Type: PEMF Duration: 7–8 h/day \times 52 weeks | Improved healing | None reported | Bronner et al. [127] |
| Femur, Tibia, radius, humerus/arthrodesis | 24 | Type: PEMF Duration: 8 h/day Settings: 75 Hz, 3.0 ± 0.5 mV | No effect | None reported | Capanna et al. [128] |
| Tibia/pseudoarthroses | 22 | Type: PEMF Duration: 8 h/day, average 5–6 ms Settings: 75 Hz, 10–20 A/cm, 180–220 V | Improved healing (90%) | Infection (three cases), protrusion of material (nine cases), Screw break (three cases) | Cebrián et al. [37] |
| Tibia/fracture | 33 | Type: PEMF Duration: 12–15/day, until healed Settings: 0.8 mT, 50 Hz | Improved healing (85%) | Infection | Colson et al. [39] |
| Tibia/fracture | 37 | Type: DC | Improved healing (100%) | None reported | Cundy and Paterson [81] |
| Tibia/non-union | 17 | Type: PEMF Duration: 20 h/day, 4–8 weeks Settings: 150–300 Gs | No effect | None reported | De Haas et al. [129] |
| Hind foot/fusion | 13 | Type: DC | Improved healing (92%) | None reported | Donley and Ward [130] |
| Humerus, tibia, femur/non-union and osteotomy | 52 | Type: PEMF Duration: 2–12 m | Improved healing (82%) | None reported | Dunn and Rush [131] |
| Cervical spine/fusion | 122 | Type: PEMF Duration: 4 h/day \times 3 m | Improved healing (83%) | None reported | Foley et al. [132] |
| Tibia/fracture | 41 | Type: DC interferential currents | No effect | Sepsis (six cases) | Fouire and Bowerbank [133] |
| Tibia, hip, radius/delayed and non-union | 12 | Type: PEMF Duration: 12 h/day \times 3 m min | No effect | None reported | Freedman [134] |
| Knee/osteoarthritis | 139 | Type: CC Duration: 6–14 h/day | Improved healing | None reported | Garland et al. [135] |
| Lumbar spine/fusion | 85 | Type: CC Duration: 24 h/day until healed or 9 m Setting: 60 kHz, current density 5 μ A root mean square/cm ² , 12 mV root mean square/cm | Improved healing (84%) | None reported | Goodwin et al. [136] |
| Tibia/non-union | 45 | Type: PEMF Duration: 12 h/day, 6–12 weeks Settings: 0.008 Weber/m ² | Improved healing (35% healed in 10 weeks and 85% in 4 ms) | Poor compliance | Gupta et al. [30] |

Table 3 (continued)

| Bone affected and/or fracture type | No. of cases | Type of ES/tim treatment | Outcome | Complications | Published article |
|--|--------------|--|-------------------------|-----------------------------|------------------------|
| Foot joint/arthropathy | 11 | Type: PEMF (combined) Duration: 0.5 h/day | Improved healing | None reported | Hanft et al. [137] |
| Hand/acute fracture | 53 | Type: PEMF Duration: continuous for 52 weeks Settings: pulse amplitude 50 mV Pulse width 5 μ s; burst width 5 ms Burst refractory period 62 ms Repeat repetition rate 15 Hz | No effect | None reported | Hannemann et al. [138] |
| Hand/acute fracture | 102 | Type: PEMF Duration: continuous max 52 weeks Settings: pulse amplitude 50 mV Pulse width 5 μ s; burst width 5 ms Burst refractory period 62 ms Repeat repetition rate 15 Hz | No effect | None reported | Hannemann et al. [139] |
| Metatarsal-foot/delayed- and non-union | 9 | Type: PEMF Duration: 8–10 h/day \times 3 m Settings: 0–20 Gs, 4.5 ms pulse bursts duration repeated at 15 Hz | Improved healing (100%) | None reported | Holmes [140] |
| Tibia/non-union | 30 | Type: PEMF Duration: 8 h/day Settings: 1–15 mV, 5 ms bursts of asymmetrical 15 Hz pulses | Improved healing (83%) | None reported | Ito and Shirai [141] |
| Radius/fracture | 18 | Type: DC (pulsed) Settings: 2 Hz, 30 μ A | Improved healing | None reported | Itoh et al. [142] |
| Lumbar spine/fusion | 17 | Type: DC and PEMF | No effect | Infection | Jenis et al. [143] |
| Tibia/fracture | 24 | Type: pulsed DC Duration: 6 m Settings: 1 Hz, 40 μ A | Improved healing (30%) | Skin reaction and infection | Jorgensen [144] |
| Tibia/fracture | 3 | Type: DC Duration: 30–60 min 3–4 \times day Settings: pulse width 300 μ s, 1–2 Hz < 20 mA | Improved healing (66%) | None reported | Kahn [145] |
| Lumbar spine/fusion | 31 | Type: DC | Improved healing (78%) | None reported | Kane [146] |
| Spine/fusion | 65 | Type: DC | Improved healing (96%) | None reported | Kucharzyk [147] |
| Radius/colles' fracture | 30 | Type: PEMF Duration: 30 m/day, 5 days/week \times 2 weeks Settings: 6 mT, 25 Hz | Improved healing | None reported | Lazovic [148] |
| Lumbar spine/fusion | 104 | Type: PEMF (combined) Duration: 30 m/day \times 9 m | Improved healing (64%) | None reported | Linovitz [149] |

Table 3 (continued)

| Bone affected and/or fracture type | No. of cases | Type of ES/tim treatment | Outcome | Complications | Published article |
|---|--------------|---|------------------------|--|----------------------------------|
| Humerus neck/fracture | 21 | Type: PEFM Duration: 30 m/day × 10 days Settings: 35 Hz, max pulse 300 W | No effect | None reported | Livesley [150] |
| Radius/non-union | 10 | Type: PEFM Duration: 104 days Settings: 2.5 Gs | Improved healing (66%) | None reported | Madronero et al. [151] |
| Tibia/osteotomy | 18 | Type: PEFM Duration: 8 h/day × 57 days Settings: pulse duration 1.3 ms, 75 Hz, 3.0 ± 0.5 mV | Improved healing | Thrombophlebitis (three cases) | Mammi et al. [152] |
| Lumbar spine/fusion | 42 | Type: PEFM Duration: 4 h/day | Improved healing (97%) | None reported | Marks [153] |
| Femur/fracture | 32 | Type: PEFM Duration: 1 h/day × 8 weeks Settings: 5–105 Hz, 0.5–2.0 mT | Improved healing (94%) | None reported | Martinez-Rondanelli et al. [154] |
| Femur head/osteonecrosis | 66 | Type: PEFM Duration: 8 h/day × 3–7 m Settings: 75 Hz, 1.3 ms pulse, 2 ± 0.5 mV | Improved healing (94%) | None reported | Massari et al. [155] |
| Mandible/fracture | 40 | Type: DC Duration: 10–14 days Settings: 10–20 µA | Improved healing | None reported | Masureik and Eriksson [156] |
| Lumbar spine/fusion | 122 | Type: DC Duration: 24 weeks min Settings: 20 µA | Improved healing (76%) | Infection (four cases) | Meril [40] |
| Tibia/delayed- and non-union | 57 | Type: PEFM | Improved healing (75%) | None reported | Meskens [157] |
| Tibia/congenital pseudarthrosis | 27 | Type: DC Duration: 6 m Settings: 20 µA | Improved healing (74%) | Infection (two cases) | Paterson and Simonis [158] |
| Humerus, ulna, radius, femur, tibia/non-union | 93 | Type: PEFM Duration: 13 weeks Settings: pulse amplitude 50 mV Pulse width 5 s at 15 Hz | Improved healing (74%) | None reported | Punt et al. [159] |
| Ankle/cystic osteochondral defect | 68 | Type: PEFM Duration: 4 h/day × 60 days Settings: 1.5 mT at 75 Hz | No effect | Temporary foot paresthesia × 2 Wound drainage × 2 | Reilingh et al. [160] |
| Lumbosacral spine/fusion | 53 | Type: DC Duration: 20.5 m Settings: 10 µA/cathode | Improved healing (96%) | None reported | Rogozinski and Rogozinski [161] |
| Foot, ankle arthrodesis/delayed union | 19 | Type: PEFM Duration: 5–27 m | Improved healing (77%) | None reported | Saltzman et al. [162] |

Table 3 (continued)

| Bone affected and/or fracture type | No. of cases | Type of ESstim treatment | Outcome | Complications | Published article |
|---|---|---|--------------------------------|---|------------------------------|
| Tibia, femur/non-union | 10 | Type: CC Duration: 6 m Setting: 5–10 V peak-to-peak sine, 60 kHz | Improved healing (60%) | Electrode allergic skin reaction (two cases) | Scott and King [163] |
| Tibia/delayed union | 20 | Type: PEMF Duration: 12 h/day × 12 weeks Settings: 200 μs with 25 μs interval, 5 T/s | Improved healing | None reported | Sharrad [28] |
| Humerus, ulna, radius, femur, tibia/non-union | 53 | Type: PEMF Duration: 12–16 h/day × 3 m Settings: 5 ms bursts, 15 Hz, 11.5 mV | Improved healing (71%) | Plate/screw loosening (eight cases) | Sharrad et al. [8] |
| Humerus, ulna, radius, femur, tibia/non-union | 31 | Type: PEMF Duration: 8 h/day × 5 m | Improved healing (77%) | None reported | Shi et al. [164] |
| Lumbar spine/fusion | 13 | Type: PEMF Duration: 8–10 h/day × 4 m Settings: 0–0.0003 T, 50 ms pulse repetition rate | Improved healing (76%) | None reported | Simmons [165] |
| Lumbar spine/fusion | 100 | Type: PEMF Duration: 2 h/day × 90 day min | Improved healing (67%) | None reported | Simmons et al. [166] |
| Metatarsal-foot/non-union | 5 | Type: PEMF Duration: 10 h/day × 24 weeks max | Improved healing | None reported | Streit et al. [167] |
| Femur head/avascular necrosis | 20 | Type: CC Duration: 24 h/day × 6 m | No effect | None reported | Steinberg et al. [168] |
| Lumbar spine/fusion | 143 | Type: DC Duration: 24 weeks Settings: 20 μA | Improved healing (91.5%) | Pain | Tejano et al. [27] |
| Ankle/cystic osteochondral defect | 68 | Type: PEMF Duration: 4 h/day × 60 days Settings: 1.5 mT at 75 Hz | Improved healing | None reported | van Bergen et al. [169] |
| Radius/fracture | 15 | Type: PEMF Duration: 8 weeks Settings: 0.00004 T at 1–1000 Hz | Improved healing | None reported | Wahlstrom and Knutsson [170] |
| Cervical spine/arthrodesis | 16 | Type: DC Duration: 26 weeks min; settings: 12 μA | Improved healing (93%) | Infection (one case) Local discomfort (four cases) | Welch et al. [171] |
| Total no. of articles | Type of bone (no.) | Type of fracture (no.) | Type of ESstim (%) | Complications (%) | Outcomes (%) |
| | Tibia Femur Spine Radius Humerus Others | Fusion Arthrodesis Delayed-/non-union Others | Osteotomy Necrosis Bone defect | PEMF DC CC | Yes No Positive Negative |
| 69 | 25 15 15 11 7 | 20 21 16 5 5 | 4 2 2 2 | 19 60 29 11 | 27 73 73 27 |

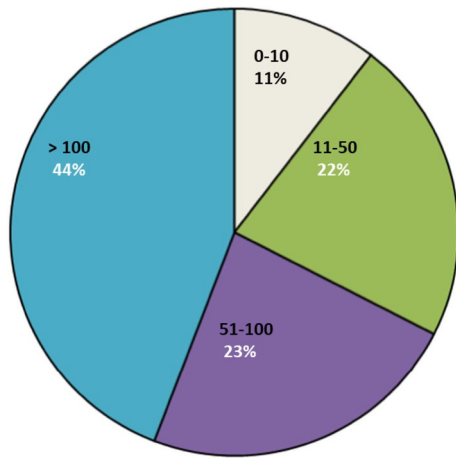


Fig. 1 How many bone surgeries do you perform per year?

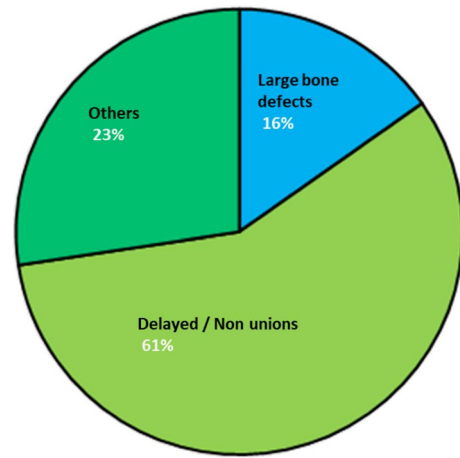


Fig. 4 In what type of bone fractures have you used an electrical stimulation device?

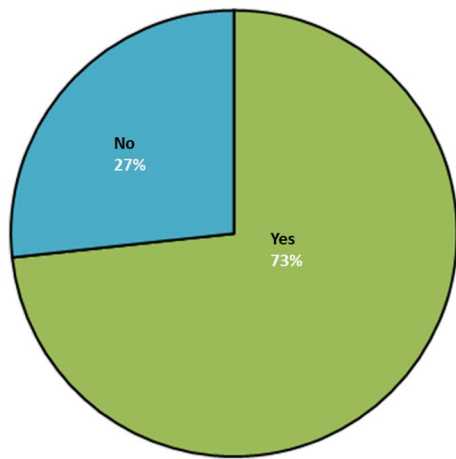


Fig. 2 Did you know that electrical stimulation has been proven to accelerate bone healing in many clinical studies?

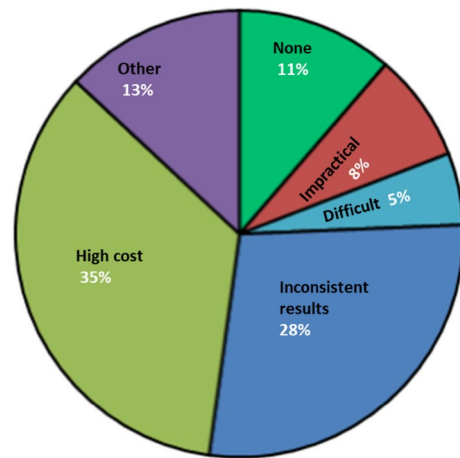


Fig. 5 What problem(s) do you see in current devices?

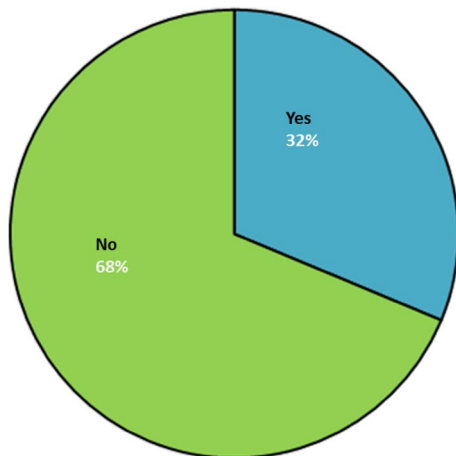


Fig. 3 Have you ever used an electrical stimulation device to treat bone fractures in your patients?

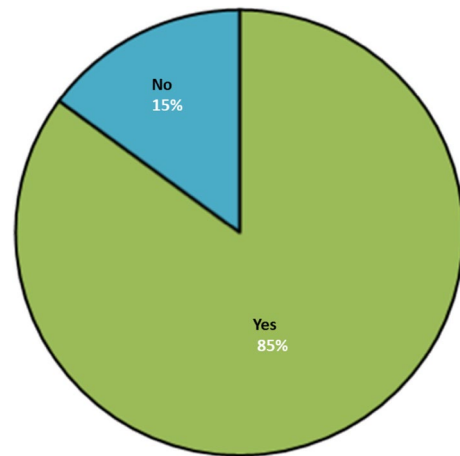


Fig. 6 If an easy-to-use electrical stimulation device to treat bone fractures were available, would you use it in your patients?

that the most commonly used method of administering EStim was PEMF while in the animal studies DC was the predominant treatment method. This preference in patients is most likely due to the fact that PEMF is administered using an external noninvasive device, whereas DC treatment requires that the EStim device be surgically implanted. While being noninvasive is a major benefit of PEMF for clinical use, patient non-compliance, associated with its use, is a major problem and is cited as one of the primary reasons for inconsistent results when using PEMF [28–30].

Another source of confusion is the dosages and regimens used. The most commonly used dosage for administering PEMF ranged between 0.3 and 6 mT, while for DC the intensity was 5–40 μ A, and for CC treatment the intensity ranged between 3 and 10 V. Half of the regimens used in the clinical studies consisted of daily EStim treatments ranging from 0.5 to 16 h/day, delivered either continuously or in interrupted intervals, the latter being for periods of 1–6 h/day. The dosages, regimens, and exposure times in the animal studies also varied widely. This great variation makes it difficult to combine the results of these studies into one or a few treatment recommendations. Another reason why it is difficult to combine the results of the different studies is because of the many different bones and fracture types studied. Of all the different bones and fracture types reported in the clinical articles by far the most common bone was the tibia and the most studied fracture types were delayed- and non-unions. In fact, this was confirmed in our survey in which orthopedic surgeons that use EStim mostly used it to treat delayed- and non-unions in their patients (Fig. 4). Again, this variation in bone and fracture types described in the literature would make it difficult to compare healing rates between, say a mandible and a tibia, or between non-unions and osteotomies, which makes it difficult to draw meaningful conclusions.

Poor fracture healing is often associated with both a lack of healing and mal-position of bone fragments. In these cases, surgeons prefer operative intervention to EStim because of the ability to restore alignment as well as facilitate fracture healing. Revision fixation and osteotomies, to correct alignment, are fraught with high rates of delayed bone healing and persistent non-unions. While EStim does not improve the position of bone fragments, it still can play an adjunctive role in the treatment of non-unions [34].

When EStim is used as an adjunct to other treatment attempts, as a last resort it can require prolonged and costly interventions. While the clinical studies we reviewed did not provide information about costs associated with EStim treatments, information available online from companies who sell clinical EStim devices indicate that the current unit cost of most EStim devices, regardless of the company (OL1000 Bone Growth Stimulator, Orthopak, EBI Bone Healing System, Physio-Stim Lite, or Exogen), is about US

\$3000. Additional costs to treat delayed unions is approximately \$24,892, that includes \$20,575 for surgery and recovery + \$4317 outpatient costs. These figures are quoted from a report published by EXOGEN [35].

Added to this, failure rates in these treatments is relatively high (17–64%) and when present can lead to additional costs [36]. Finally, the costs associated with using EStim devices are usually not reimbursed, thus further reducing the incentive to use this treatment option. While comparing the costs of EStim to other treatments used in problematic fractures is beyond the scope of this paper, from the responses we received in our survey it is clear that high costs is an important factor for surgeons in their decision whether or not to use EStim. Of the drawbacks associated with using EStim, the greatest number of surgeons surveyed cited high costs as being the main problem with using EStim in their fracture patients (Fig. 5).

The second most cited problem with EStim was inconsistent results. While the questions in our survey did not ask about the specific cause of the inconsistent results associated with using EStim, from our literature review we were able to identify some possible causes. In the clinical papers a few different device-related problems were cited that could cause “inconsistent results”. These included “damaged” or “disconnected” implanted stimulators, misplacement of hardware, and migration of the EStim device’s electrode leads that can occur due to muscle movement or insufficient flexibility of the muscles [9, 28, 37]. Eight and five percentage of surgeons surveyed indicated that the problems they encounter using EStim were associated to the devices used to administer the treatment, choosing “impractical”, and “difficult to use”, respectively, to describe their experience. In a white paper generated by industry that compares costs associated with the use of five different EStim bone stimulators the authors write that using these devices the “probability of failure” ranges between 17 and 64% [36]. While the exact causes of failure are not specified in this paper these high failure rates could certainly cause inconsistent results.

Of the 69 clinical studies we reviewed, 19 reported complications experienced during treatment with EStim. In these 19 studies, the most common type of complication experienced was skin irritation and infections, when using the externally applied PEMF device, and infections at the fracture line when using the implanted DC device [24–27]. Other types of complications experienced with EStim treatment were pain [27], dislocation of the device [28], failure of the device [29] and poor patient compliance [30]. The above-mentioned complications and particularly patient non-compliance could be other causes of the inconsistent results surgeons cited in our survey. Existing external PEMF units are cumbersome and require many hours of treatment per day over months, which interferes with activities of daily living, causing decrease compliance. If a patient does not (or

is not able to) utilize the PEMF EStim device in the manner prescribed then the beneficial effects are diminished. Smaller units are available and only require 30-min treatments, however, they require very precise fitting to encompass the fracture site within the small field which also decreases the effectively applied dose and clinical efficacy. Although 73% of clinical studies demonstrate a benefit to EStim, the magnitude and consistency of the effect are less than reported in animal studies. Patient compliance is much lower in clinical studies than in animal studies. We believe that problems with compliance account for the large gap in the results reported in the clinical versus animal studies.

In a study we reviewed, Simmons et al. compared PEMF (where patient compliance is required) to DC EStim (where compliance is not an issue since the device is implanted), and found that spinal fusion rates in the former were lower than in the latter [41], attributing this difference to patient non-compliance. In another study non-compliance was cited as a possible reason for EStim-treated patients having the same spinal fusion rates as non-treated controls [42]. The above problems, “high cost”, “inconsistent results”, and “impractical/difficult to use” go a long way toward answering our original question, why EStim-based fracture treatments have not gained more acceptance in the orthopedic community.

When comparing EStim to other adjunct treatments used to treat delayed healing or non-unions, Ebrahim et al. compared EStim with low intensity-pulsed ultrasound and found no significant difference [43]. Kertzman et al. used radial extracorporeal shock wave therapy to treat fracture non-unions of superficial bones and found that 70% of tibia non-unions healed within 6 months suggesting that this approach is on par with EStim [44]. Similarly, in a recent study by Putnam et al. non-unions in 26 patients using surgical volar plate fixation and cancellous grafting, they found 82% healed by 12 weeks [45]. With this, one can reasonably assume that rates of success with these different procedures are similar to those reported with EStim.

The present study had several drawbacks, the greatest being difficulty making sense of the large variation in the methods reported in the different studies we reviewed. In both the animal and clinical studies, the bones and fracture types studied, the EStim method/dosages/regimens, and the methods used to report outcomes differed greatly. This made it difficult to combine these various parameters into well-founded treatment recommendations. Another weakness in this study was positive publication bias. All the articles we reviewed were published, and since studies that found EStim to be effective are more likely to be written up, submitted, and accepted for publication, our review did not include unpublished studies with negative results. Another shortfall is related to the questions we used in our survey. We did not test these questions for

validity or reproducibility prior to sending them out. Had this been done perhaps we could have improved the quality of the answers we received. Finally, in the survey we could have included more specific questions that might have provided answers to other important questions such as the specific causes of the problem’s respondents encountered with EStim treatment. While it would have been nice to get more information with more detailed questions, we decided to have few and simple questions thinking that this would help maximize the response rate in this first study.

Conclusion

Most of the orthopedic surgeons we surveyed were aware of EStim and its positive outcomes in fracture treatment. These positive outcomes were confirmed in the literature we reviewed, in which both preclinical animal and clinical studies reported positive overall outcomes using EStim to treat bone fractures. Despite the awareness and positive impression our respondents had about EStim only a fraction actually use it in their fracture patients. The reason for this discrepancy could be problems such as, confusion in the literature, due to the great variation in methods reported, and the inconsistent results associated with this treatment approach. On the positive side, when asked “If an easy-to-use electrical stimulation device to treat bone fractures were available, would you use it in your patients?”, 85% of the surgeons surveyed responded “Yes”. This suggests that despite the problems, given an easy-to-use method for administering EStim, surgeons are open to using this treatment approach. An improved delivery system for EStim could overcome the compliance problem, markedly increase the clinical efficacy and make EStim an accepted form of treatment of non-unions and acute fractures associated with poor healing.

Funding This study was supported in part by the Friedrichsheim Foundation (Stiftung Friedrichsheim) based in Frankfurt/Main, Germany, and the Chinese Scholarship Council (CSC).

Compliance with ethical standards

Conflict of interest The authors have no conflicts of interest to declare.

References

1. Garrat AC. *Electrophysiology and electrotherapeutics*. Boston: Ticknor and Fields; 1860.
2. Kuzyk PR, Schemitsch EH. The science of electrical stimulation therapy for fracture healing. *Indian J Orthop*. 2009;43(2):127–31. <https://doi.org/10.4103/0019-5413.50846>.

3. Chalidis B, Sachinis N, Assiotis A, Maccauro G, Graziani F. Stimulation of bone formation and fracture healing with pulsed electromagnetic fields: biologic responses and clinical implications. *Int J Immunopathol Pharmacol*. 2011;24:17–20.
4. Aaron RK, Boyan BD, Ciombor DM, Schwartz Z, Simon BJ. Stimulation of growth factor synthesis by electric and electromagnetic fields [review]. *Clin Orthop*. 2004;419:30–7.
5. Simonis RB, Parnell EJ, Ray PS, Peacock JL. Electrical treatment of tibial non-union: a prospective, randomised, double-blind trial. *Injury*. 2003;34:357–62.
6. Andersen T, Christensen FB, Ernst C, Fruensgaard S, Østergaard J, Andersen JL, et al. The effect of electrical stimulation on lumbar spinal fusion in older patients: a randomized, controlled, multi-center trial: part 1: functional outcome. *Spine*. 2009;34:2241–7.
7. Steinberg ME, Brighton CT, Hayken GD, Tooze SE, Steinberg DR. Early results in the treatment of avascular necrosis of the femoral head with electrical stimulation. *Orthop Clin N Am*. 1984;15:163–75.
8. Sharrard WJ, Sutcliffe ML, Robson MJ, Maceachern AG. The treatment of fibrous non-union of fractures by pulsing electromagnetic stimulation. *J Bone Jt Surg Br*. 1982;64:189–93.
9. Brighton C, Shaman P, Heppenstall R. Tibial nonunion treated with direct current, capacitive coupling, or bone graft. *Clin Orthop*. 1995;321:223–34.
10. Borsalino G, Bagnacani M, Bettati E, et al. Electrical stimulation of human femoral intertrochanteric osteotomies. *Clin Orthop*. 1988;237:256–63.
11. Bassett CA, Mitchell SN, Schink MM. Treatment of therapeutically resistant non-unions with bone grafts and pulsing electromagnetic fields. *J Bone Jt Surg Am*. 1982;64:1214–20.
12. Steinberg ME, Brighton CT, Corces A, Hayken GD, Steinberg DR, Stafford B, et al. Osteonecrosis of the femoral head. Results of core decompression and grafting with and without electrical stimulation. *Clin Orthop Relat Res*. 1989;249:199–208.
13. Tai G, Tai M, Zhao M. Electrically stimulated cell migration and its contribution to wound healing. *Burns Trauma*. 2018;6:20. <https://doi.org/10.1186/s41038-018-0123-2>.
14. Yuan X, Arkonac DE, Chao PG, Vunjak-Novakovic G. Electrical stimulation enhances cell migration and integrative repair in the meniscus. *Sci Rep*. 2014;4:3674. <https://doi.org/10.1038/srep03674>.
15. Ercan B, Webster TJ. Greater osteoblast proliferation on anodized nanotubular titanium upon electrical stimulation. *Int J Nanomed*. 2008;3(4):477–85.
16. Guo BS, Cheung KK, Yeung SS, Zhang BT, Yeung EW. Electrical stimulation influences satellite cell proliferation and apoptosis in unloading-induced muscle atrophy in mice. *PLoS One*. 2012;7(1):e30348. <https://doi.org/10.1371/journal.pone.0030348>.
17. Serena E, Figallo E, Tandon N, Cannizzaro C, Gerech S, Elvasore N, et al. Electrical stimulation of human embryonic stem cells: cardiac differentiation and the generation of reactive oxygen species. *Exp Cell Res*. 2009;315(20):3611–9. <https://doi.org/10.1016/j.yexcr.2009.08.015>.
18. Yamada A, Gaja N, Ohya S, Muraki K, Narita H, Ohwada T, et al. Usefulness and limitation of DiBAC4(3), a voltage-sensitive fluorescent dye, for the measurement of membrane potentials regulated by recombinant large conductance Ca^{2+} -activated K^{+} channels in HEK293 cells. *Jpn J Pharmacol*. 2001;86(3):342–50.
19. Eischen-Loges M, Oliveira KMC, Bhavsar MB, Barker JH, Leppik L. Pretreating mesenchymal stem cells with electrical stimulation causes sustained long-lasting pro-osteogenic effects. *PeerJ*. 2018;6:e4959. <https://doi.org/10.7717/peerj.4959>.
20. Mobini S, Leppik L, Parameswaran VT, Barker JH. In vitro effect of direct current electrical stimulation on rat mesenchymal stem cells. *PeerJ*. 2017;12(5):e2821. <https://doi.org/10.7717/peerj.2821>.
21. Behari J. Effect of electrical stimulation in mineralization and collagen enrichment of osteoporotic rat bones. In: 2008 International conference on recent advances in microwave theory and applications 2008.
22. Durigan JLQ, Peviani SM, Delfino GB, De Souza Jose RJ, Parra T, Salvini TF. Neuromuscular electrical stimulation induces beneficial adaptations in the extracellular matrix of quadriceps muscle after anterior cruciate ligament transection of rats. *Am J Phys Med Rehabil*. 2014;93(11):948–61. <https://doi.org/10.1097/PHM.000000000000110>.
23. George PM, Bliss TM, Hua T, Lee A, Oh B, Levinson A, et al. Electrical preconditioning of stem cells with a conductive polymer scaffold enhances stroke recovery. *Biomaterials*. 2017;142:31–40. <https://doi.org/10.1016/j.biomaterials>.
24. Leppik LP, Froemel D, Slavici A, Ovidia ZN, Hudak L, Henrich D, et al. Effects of electrical stimulation on rat limb regeneration, a new look at an old model. *Sci Rep*. 2015;17(5):18353. <https://doi.org/10.1038/srep18353>.
25. Leppik L, Zhihua H, Mobini S, Parameswaran VT, Eischen-Loges M, Slavici A, et al. Combining electrical stimulation and tissue engineering to treat large bone defects in a rat model. *Sci Rep*. 2018;8(1):6307. <https://doi.org/10.1038/s41598-018-24892-0>.
26. Brochet F, Weber J. LinkedIn Corporation. Harvard Business School Case 112–006; 2012.
27. Tejano N, Puno R, Ignacio JM. The use of implantable direct current stimulation in multilevel spinal fusion without instrumentation. *Spine*. 1996;21(16):1904–8. <https://doi.org/10.1097/00007632-199608150-00015>.
28. Sharrard WJW. A double-blind trial of pulsed electromagnetic fields for delayed union of tibial fractures. *J Bone Jt Surg Br*. 1990;72:347–55.
29. Brighton CT. Treatment of non-union of the tibia with constant direct current. *J Trauma*. 1981;21:189–95.
30. Gupta AK, Srivastava KP, Avasthi S. Pulsed electromagnetic stimulation in nonunion of tibial diaphyseal fractures. *Indian J Orthop*. 2009;43(2):156–60. <https://doi.org/10.4103/0019-5413.50850>.
31. Spadaro JA. Electrically stimulated bone growth in animals and man. Review of the literature. *Clin Orthop Relat Res*. 1977;122:325–32.
32. Barker AT, Dixon RA, Sharrard WJW, Sutcliffe ML. Pulsed magnetic field therapy for tibial non-union. Interim results of a double-blind trial. *Lancet*. 1984;1:994–6.
33. Brighton C, Pollack S. Treatment of recalcitrant non-unions with a capacitively coupled electrical field. *J Bone Joint Surg*. 1985;67A:577–85.
34. Kooistra BW, Jain A, Hanson BP. Electrical stimulation: nonunions. *Indian J Orthop*. 2009;43(2):149–55.
35. EXOGEN. EXOGEN® Bone healing system shown to be most cost-effective bone stimulator. 2005.
36. Schultz M, Oremus M, Whitman C, Conway J. Cost-effectiveness of bone stimulators in the conservative treatment of stable nonunion fractures. *Value Health*. 2004;7:723 (**International Society for Pharmacoeconomics and Outcomes Research (ISPOR)**).
37. Cebrián JL, Gallego P, Francés A, Sánchez P, Manrique E, Marco F, et al. Comparative study of the use of electromagnetic fields in patients with pseudoarthrosis of tibia treated by intramedullary nailing. *Int Orthop*. 2010;34(3):437–40. <https://doi.org/10.1007/s00264-009-0806-1>.
38. Abdelrahim A, Hassanein HR, Dahaba M. Effect of pulsed electromagnetic field on healing of mandibular fracture: a preliminary clinical study. *J Oral Maxillofac Surg*. 2011;69(6):1708–17. <https://doi.org/10.1016/j.joms.2010.10.013>.

39. Colson DJ, Browett JP, Fiddian NJ, Watson B. Treatment of delayed- and non-union of fractures using pulsed electromagnetic fields. *J Biomed Eng.* 1988;10:301–4.
40. Meril AJ. Direct current stimulation of allograft in anterior and posterior lumbar interbody fusions. *Spine.* 1994;19:2393–8.
41. Simmons JW, Hayes MA, Christensen DK, Dwyer AP, Koullis CS, Kimmich SJ. The effect of postoperative pulsing electromagnetic fields on lumbar fusion: an open trial phase study. Quebec, Canada: Presented at the North American Spine Society; 1989.
42. Lee K. Clinical investigation of the spinal stem system open trial phase: pseudarthrosis stratum. Las Vegas, Nevada: Presented at the annual meeting of the American Academy of Orthopaedic Surgeons; 1989.
43. Ebrahim S, Mollon B, Bance S, Busse JW, Bhandari M. Low-intensity pulsed ultrasonography versus electrical stimulation for fracture healing: a systematic review and network meta-analysis. *Can J Surg.* 2014;57(3):E105–18.
44. Kertzman P, Császár NBM, Furia JP, Schmitz C. Radial extracorporeal shock wave therapy is efficient and safe in the treatment of fracture nonunions of superficial bones: a retrospective case series. *J Orthop Surg Res.* 2017;12(1):164. <https://doi.org/10.1186/s13018-017-0667-z>.
45. Putnam JG, Mitchell SM, DiGiovanni RM, Stockwell EL, Edwards SG. Outcomes of unstable scaphoid nonunion with segmental defect treated with plate fixation and autogenous cancellous graft. *J Hand Surg Am.* 2019;44(2):160.e1–7. <https://doi.org/10.1016/j.jhsa.2018.05.023>.
46. Aydin N, Bezer M. The effect of an intramedullary implant with a static magnetic field on the healing of the osteotomised rabbit femur. *Int Orthop.* 2011;35(1):135–41. <https://doi.org/10.1007/s00264-009-0932-9>.
47. Barak S, Neuman M, Iezzi G, Piattelli A, Perrotti V, Gabet Y. A new device for improving dental implants anchorage: a histological and micro-computed tomography study in the rabbit. *Clin Oral Implant Res.* 2016;27(8):935–42. <https://doi.org/10.1111/clr.12661>.
48. Buzza EP, Shibli JA, Barbeiro RH, Barbosa JR. Effects of electromagnetic field on bone healing around commercially pure titanium surface: histologic and mechanical study in rabbits. *Implant Dent.* 2003;12:182–7.
49. Fredericks DC, Piehl DJ, Baker JT, Abbott J, Nepola JV. Effects of pulsed electromagnetic field stimulation on distraction osteogenesis in the rabbit tibial leg lengthening model. *J Pediatr Orthop.* 2003;23:478–83.
50. France JC, Norman TL, Santrock RD, McGrath B, Simon BJ. The efficacy of direct current stimulation for lumbar intertransverse process fusions in an animal model. *Spine.* 2001;26:1002–8.
51. Gilotra M, Griffith C, Schiavone J, Nimmagadda N, Nouveau J, Ludwig SC. Capacitive coupling reduces instrumentation-related infection in rabbit spines: a pilot study. *Clin Orthop Relat Res.* 2012;470(6):1646–51. <https://doi.org/10.1007/s11999-011-2231-1>.
52. Hu J, Qu J, Xu D, Zhang T, Qin L, Lu H. Combined application of low-intensity pulsed ultrasound and functional electrical stimulation accelerates bone-tendon junction healing in a rabbit model. *J Orthop Res.* 2014;32(2):204–9. <https://doi.org/10.1002/jor.22505>.
53. Kim J, Yang HJ, Cho TH, Lee SE, Park YD, Kim HM, et al. Enhanced regeneration of rabbit mandibular defects through a combined treatment of electrical stimulation and rhBMP-2 application. *Med Biol Eng Comput.* 2013;51(12):1339–48. <https://doi.org/10.1007/s11517-013-1106-x>.
54. Matsumoto H, Ochi M, Abiko Y, Hirose Y, Kaku T, Sakaguchi K. Pulsed electromagnetic fields promote bone formation around dental implants inserted into the femur of rabbits. *Clin Oral Implant Res.* 2000;11(4):354–60.
55. Ottani V, Raspanti M, Martini D, Tretola G, Ruggeri A, Franchi M, et al. Electromagnetic stimulation on the bone growth using backscattered electron imaging. *Micron.* 2002;33:121–5.
56. Rubinacci A, Black J, Brighton CT, Friedenber ZB. Changes in bioelectric potentials on bone associated with direct current stimulation of osteogenesis. *J Orthop Res.* 1988;6:335–45.
57. Shafer DM, Rogerson K, Norton L, Bennett J. The effect of electrical perturbation on osseointegration of titanium dental implants. *J Oral Maxillofac Surg.* 1995;53:1063–8.
58. Shimizu E, Matsuda-Honjyo Y, Samoto H, Saito R, Nakajima Y, Nakayama Y, et al. Static magnetic fields-induced bone sialoprotein (BSP) expression is mediated through FGF2 response element and pituitary-specific transcription factor-1 motif. *J Cell Biochem.* 2004;91:1183–96.
59. Smith R, Nagel D. Effects of pulsing electromagnetic fields on bone growth and articular cartilage. *Clin Orthop.* 1983;181:277–82.
60. Taylor BC, French BG, Fowler TT, Russell J, Poka A. Induced membrane technique for reconstruction to manage bone loss. *J Am Acad Orthop Surg.* 2012;20:142–50.
61. Veronesi F, Cadossi M, Giavaresi G, Martini L, Setti S, Buda R, et al. Pulsed electromagnetic fields combined with a collagenous scaffold and bone marrow concentrate enhance osteochondral regeneration: an in vivo study. *BMC Musculoskelet Disord.* 2015;2(16):233. <https://doi.org/10.1186/s12891-015-0683-2>.
62. Yonemori K, Matsunaga S, Ishidou Y, Maeda S, Yoshida H. Early effects of electrical stimulation on osteogenesis. *Bone.* 1996;19:173–80.
63. Zimmerman M, Parsons JR, Alexander H, Weiss AB. The electrical stimulation of bone using a filamentous carbon cathode. *J Biomed Mater Res.* 1984;18:927–38.
64. Berry JL, Geiger JM, Moran JM, Skraba JS, Greenwald AS. Use of tricalcium phosphate or electrical-stimulation to enhance the bone porous implant interface. *J Biomed Mater Res.* 1986;20:65–77.
65. Bins-Ely LM, Cordero EB, Souza JCM, Teughels W, Benfatti CAM, Magini RS. In vivo electrical application on titanium implants stimulating bone formation. *J Periodontol Res.* 2017;52(3):479–84. <https://doi.org/10.1111/jre.12413>.
66. Branham GB, Triplett RG, Yeandle S, Vieras F. The effect of electrical current on the healing of mandibular freeze-dried bone allografts in dogs. *J Oral Maxillofac Surg.* 1985;43(6):403–7.
67. Chakkalakal DA, Lippiello L, Shindell RL, Connolly JF. Electrophysiology of direct current stimulation of fracture healing in canine radius. *IEEE Trans Biomed Eng.* 1990;37:1048–58.
68. Colella SM, Miller AG, Stang RG, Stoebe TG, Spengler DM. Fixation of porous titanium implants in cortical bone enhanced by electrical stimulation. *J Biomed Mater Res.* 1981;15:37–46.
69. Connolly JF, Henry H, Jardon J. The Electrical Enhancement of Periosteal Proliferation in Normal and Delayed Fracture Healing. *Clin Orthop.* 1977;124:97–105.
70. Dejardin LM, Kahanovitz N, Arnoczky SP, Simon BJ. The effect of varied electrical current densities on lumbar spinal fusions in dogs. *Spine J.* 2001;1:341–7.
71. Doyle ND. Rehabilitation of fractures in small animals: maximize outcomes, minimize complications. *Clin Tech Small Anim Pract.* 2004;19:180–91.
72. Rodriguez Fuentes AE, Marcondes de Souza JP, Valeri V, Mascarenhas S. Experimental model of electric stimulation of pseudarthrosis healing. *Clin Orthop.* 1984;183:267–75.
73. Inoue N, Ohnishi I, Chen D, Deitz LW, Schwardt JD, Chao EYS. Effect of pulsed electromagnetic fields (PEMF) on late-phase osteotomy gap healing in a canine tibial model. *J Orthop Res.* 2002;20:1106–14.
74. Jacobs JD, Norton LA. Electrical stimulation of osteogenesis in periodontal defects. *Clin Orthop.* 1977;124:41–52.

75. Jacobs RR, Luethi U, Dueland RT, Perren SM. Electrical stimulation of experimental nonunions. *Clin Orthop Relat Res.* 1981;161:146–53.
76. Kahanovitz N, Arnoczky S, Nemzek J, Shores A. The effect of EMF pulsing on posterior lumbar spinal fusion in dogs. *Spine.* 1994;19:705–9.
77. Lindsey RW, Grobman J, Leggon RE, Panjabi M, Friedlaender GE. Effects of bone graft and electrical stimulation on the strength of healing bony defects in dogs. *Clin Orthop.* 1987;222:275–80.
78. Modarresi J, Aghili H, Karandish M, Jalali B, Zahir ST. Effect of direct electric current on parietal bone osteogenesis. *J Craniofac Surg.* 2012;23(6):1607–9. <https://doi.org/10.1097/SCS.0b013e3182575423>.
79. Ortman LF, Casey DM, Deers M. Bioelectric stimulation and residual ridge resorption. *J Prosthet Dent.* 1992;67:67–71.
80. Dev MED, Org ART, Ingrowth T, Recum V, Al PET. ABSTRACT The effect of electrical stimulation on the interfacial strength of the porous polymethylmethacrylate implant/oral tissue union and the amount. Department of Interdisciplinary Studies, College of Engineering Clemson University Clemson, 1978;6:291–303.
81. Cundy PJ, Paterson DC. A ten year review of treatment of delayed union and nonunion with an implanted bone growth stimulator. *Clin Orthop Relat Res.* 1988;259:216–22.
82. Paterson DC, Hillier TM, Carter RF, Ludbrook J, Maxwell GM, Savage JP. Experimental delayed union of the dog tibia and its use in assessing the effect of an electrical bone growth stimulator. *Clin Orthop.* 1977;128:340–50.
83. Paterson DC, Carter RF, Tilbury RF, Ludbrook J, Savage JP. The effects of varying current levels of electrical stimulation. *Clin Ortho Relat Res.* 1982;169:303–12.
84. Pepper JR, Herbert MA, Anderson JR, Bobeck WP. Effect of capacitive coupled electrical stimulation on regenerate bone. *J Orthop Res.* 1996;14:296–302.
85. Schutzer SF, Jasty M, Bragdon CR, Harrigan TP, Harris WH. A double-blind study on the effects of a capacitively coupled electrical field on bone ingrowth into porous-surfaced canine total hip prosthesis. *Clin Orthop Rel Res.* 1990;260:297–304.
86. Shayesteh YS, Eslami B, Dehghan MM, Vaziri H, Alikhassi M, Mangoli A, et al. The effect of a constant electrical field on osseointegration after immediate implantation in dog mandibles: a preliminary study: basic science research. *J Prosthodont.* 2007;16:337–42.
87. Shokry M. Preliminary study on the use of a silver oxide watch battery (1.5 V) for electrical enhancement of bone healing. *Vet Res Commun.* 1985;9:245–50.
88. Srivastava KP, Lahiri V, Khare A, Chandra H. Histomorphologic evidence of fracture healing after direct electrical stimulation in dogs. *J Trauma.* 1982;22(9):785–6.
89. Atalay Y, Gunes N, Guner MD, Akpolat V, Celik MS, Guner R. Pentoxifylline and electromagnetic field improved bone fracture healing in rats. *Drug Des Dev Ther.* 2015;9(9):5195–201. <https://doi.org/10.2147/DDDT.S89669>.
90. Brighton CT, Tadduni GT, Goll SR, Pollack SR. Treatment of denervation/diuse osteoporosis in the rat with a capacitively coupled electrical signal: effects on bone formation and bone resorption. *J Orthop Res.* 1988;6:676–84.
91. Giannunzio GA, Speerli RC, Guglielmotti MB. Electrical field effect on peri-implant osteogenesis: a histologic and histomorphometric study. *Implant Dent.* 2008;17:118–26.
92. Guizzardi S, Silvestre M, Govoni P, Scandroglio R. Pulsed electromagnetic field stimulation on posterior spinal fusions: a histological study in rats. *J Spinal Disord.* 1994;7:36–40.
93. Van Der Jagt OP, Van Der Linden JC, Waarsing JH, Verhaar JAN, Weinans H. Systemic treatment with pulsed electromagnetic fields do not affect bone microarchitecture in osteoporotic rats. *Int Orthop.* 2012;36(7):1501–6. <https://doi.org/10.1007/s00264-011-1471-8>.
94. Lirani-Galvão APR, Bergamaschi CT, Silva OL, Lazaretti-Castro M. Electrical field stimulation improves bone mineral density in ovariectomized rats. *Braz J Med Biol Res.* 2006;39:1501–5.
95. Manjhi J, Mathur R, Behari J. Effect of low level capacitive-coupled pulsed electric field stimulation on mineral profile of weight-bearing bones in ovariectomized rats. *J Biomed Mater Res B Appl Biomater.* 2010;92(1):189–95. <https://doi.org/10.1002/jbm.b.31505>.
96. Marino AA, Cullen JM, Reichmanis M, Becker RO. Fracture healing in rats exposed to extremely low frequency electric fields. *Clin Orthop* 1979;145:239–44.
97. Medalha CC, Amorim BO, Ferreira JM, Oliveira P, Pereira RMR, Tim C, et al. Comparison of the effects of electrical field stimulation and low-level laser therapy on bone loss in spinal cord-injured rats. *Photomed Laser Surg.* 2010;28(5):669–74. <https://doi.org/10.1089/pho.2009.2691>.
98. Nakajima M, Inoue M, Hojo T, Inoue N, Tanaka K, Takatori R, et al. Effect of electroacupuncture on the healing process of tibia fracture in a rat model: a randomised controlled trial. *Acupunct Med.* 2010;28:140–3.
99. Puricelli E, Dutra NB, Ponzoni D. Histological evaluation of the influence of magnetic field application in autogenous bone grafts in rats. *Head Face Med.* 2009;5:1. <https://doi.org/10.1186/1746-160X-5-1>.
100. Shen WW, Zhao JH. Pulsed electromagnetic fields stimulation affects BMD and local factor production of rats with disuse osteoporosis. *Bioelectromagnetics.* 2010;31(2):113–9. <https://doi.org/10.1002/bem.20535>.
101. Spadaro JA, Becker RO. Function of implanted cathodes in electrode-induced bone growth. *Med Biol Eng Comput.* 1979;17:769–75.
102. Takano-Yamamoto T, Kawakami M, Sakuda M. Effect of a pulsing electromagnetic field on demineralized bone-matrix-induced bone formation in a bony defect in the premaxilla of rats. *J Dent Res.* 1992;71:1920–5.
103. Tomofuji T, Ekuni D, Azuma T, Irie K, Endo Y, Kasuyama K, et al. Effects of electrical stimulation on periodontal tissue remodeling in rats. *J Periodontol Res.* 2013;48(2):177–83. <https://doi.org/10.1111/j.1600-0765.2012.01518.x>.
104. Uysal T, Amasyali M, Olmez H, Karslioglu Y, Gunhan O. Stimulation of bone formation by direct electrical current in an orthopedically expanded suture in the rat. *Korean J Orthod.* 2010;40:106–14.
105. Yang BY, Huang TC, Chen YS, Yao CH. Reconstructive effects of percutaneous electrical stimulation combined with GGT composite on large bone defect in rats. *Evid Based Complement Altern Med.* 2013. <https://doi.org/10.1155/2013/607201>.
106. Yu K, Yoon YS, Jeon J. The effect of electrical stimulation combined with foam dressing on ulcer healing in rats with spinal cord injury. *Adv Skin Wound Care.* 2015;28(11):495–502. <https://doi.org/10.1097/01.ASW.0000470553.85257.84>.
107. Zamarioli A, Battaglini RA, Morse LR, Sudhakar S, Maranhão DAC, Okubo R, et al. Standing frame and electrical stimulation therapies partially preserve bone strength in a rodent model of acute spinal cord injury. *Am J Phys Med Rehabil.* 2013;92(5):402–10. <https://doi.org/10.1097/PHM.0b013e318287697c>.
108. Benazzo F, Cadossi M, Cavani F, Fini M, Giavaresi G, Setti S, et al. Cartilage repair with osteochondral autografts in sheep: effect of biophysical stimulation with pulsed electromagnetic fields. *J Orthop Res.* 2008;26(5):631–42. <https://doi.org/10.1002/jor.20530>.
109. Dergin G, Aktas M, Gürsoy B, Kürkçü M, Devecioğlu Y, Benlidayı E. Direct current electric stimulation in implant

- osseointegration: an experimental animal study with sheep. *J Oral Implantol*. 2013;39(6):671–9. <https://doi.org/10.1563/AAID-JOI-D-10-00172>.
110. Flouty OE, Oya H, Kawasaki H, Reddy CG, Fredericks DC, Gibson-Corley KN, et al. Intracranial somatosensory responses with direct spinal cord stimulation in anesthetized sheep. *PLoS One*. 2013;8(2):e56266. <https://doi.org/10.1371/journal.pone.0056266>.
 111. El-Hakim IE, Azim AM, El-Hassan MF, Maree SM. Preliminary investigation into the effects of electrical stimulation on mandibular distraction osteogenesis in goats. *Int J Oral Maxillofac Surg*. 2004;33(1):42–7.
 112. Law HT, Annan I, McCarthy ID, Hughes SP, Stead AC, Camburn MA, et al. The effect of induced electric currents on bone after experimental osteotomy in sheep. *J Bone Jt Surg Br*. 1985;67:463–9.
 113. Muttini A, Abate M, Bernabò N, Cavani F, Mingozzi R, Tosi U, et al. Effect of electric current stimulation in combination with external fixator on bone healing in a sheep fracture model. *Vet Ital*. 2014;50(4):249–57. <https://doi.org/10.12834/VetIt/271.963>.
 114. Toth JM, Seim HB, Schwardt JD, Humphrey WB, Wallskog JA, Turner AS. Direct current electrical stimulation increases the fusion rate of spinal fusion cages. *Spine*. 2000;25:2580–7.
 115. Canè V, Botti P, Farneti D, Soana S. Electromagnetic stimulation of bone repair: a histomorphometric study. *J Orthop Res*. 1991;9:908–17.
 116. Kold SE, Hickman J. Preliminary study of quantitative aspects and the effect of pulsed electromagnetic field treatment on the incorporation of equine cancellous bone graft. *Equine Vet J*. 1987;19(2):120–4.
 117. Sanders-Shamis M, Bramlage LR, Weisbrode SE, Gabel AA. A preliminary investigation of the effect of selected electromagnetic field devices on healing of cannon bone osteotomies in horses. *Equine Vet J*. 1989;21:201–5.
 118. Abeed RI, Naseer M, Abel EW. Capacitively coupled electrical stimulation treatment: results from patients with failed long bone fracture unions. *J Orthop Trauma*. 1998;12:510–3.
 119. Adie S, Harris I, Naylor J. Pulsed electromagnetic field stimulation for acute tibial shaft fractures: a multicenter, double-blind, randomized trial. *J Bone Jt Surg Am*. 2011;93(17):1569–76. <https://doi.org/10.2106/JBJS.J.00869>.
 120. Andersen T, Christensen FB, Egund N, Ernst C, Fruensgaard S, Ostergaard J, et al. The effect of electrical stimulation on lumbar spinal fusion in older patients: a randomized, controlled, multicenter trial: part 2: fusion rates. *Spine*. 2009;34:2248–53.
 121. Andersen T, Christensen FB, Langdahl BL, Ernst C, Fruensgaard S, Østergaard J, et al. Fusion mass bone quality after uninstrumented spinal fusion in older patients. *Eur Spine J*. 2010;19(12):2200–8. <https://doi.org/10.1007/s00586-010-1373-2>.
 122. Assiotis A, Sachinis NP, Chalidis BE. Pulsed electromagnetic fields for the treatment of tibial delayed unions and nonunions A prospective clinical study and review of the literature. *J Orthop Surg Res*. 2012;7:24. <https://doi.org/10.1186/1749-799x-7-24>.
 123. Bassett CA, Mitchell SN, Gaston SR. Treatment of ununited tibial diaphyseal fractures with pulsing electromagnetic fields. *J Bone Jt Surg Am*. 1981;63:511–23.
 124. Beck BR, Matheson GO, Bergman G, Norling T, Fredericson M, Hoffman AR, et al. Do capacitively coupled electric fields accelerate tibial stress fracture healing? A randomized controlled trial. *Am J Sports Med*. 2008;36(3):545–53.
 125. Benazzo F, Mosconi M, Beccarisi G, Galli U. Use of capacitive coupled electric fields in stress fractures in athletes. *Clin Orthop Relat Res*. 1995;310:145–9.
 126. Boyette MY, Herrera-Soto JA. Treatment of delayed and nonunited fractures and osteotomies with pulsed electromagnetic field in children and adolescents. *Orthopedics*. 2012;35(7):e1051–5. <https://doi.org/10.3928/01477447-20120621-20>.
 127. Bronner S, Novella T, Becica L. Management of a delayed-union sesamoid fracture in a dancer. *J Orthop Sports Phys Ther*. 2007;37:529–40.
 128. Capanna R, Donati D, Masetti C, Manfrini M, Panozzo A, Cadossi R, et al. Effect of electromagnetic fields on patients undergoing massive bone graft following bone tumor resection. A double blind study. *Clin Orthop Rel Res*. 1994;306:213–21.
 129. de Haas WG, Watson J, Morrison DM. Non-invasive treatment of ununited fractures of the tibia using electrical stimulation. *J Bone Jt Surg Br*. 1980;62:465–70.
 130. Donley BG, Ward DM. Implantable electrical stimulation in high-risk hindfoot fusions. *Foot Ankle Int*. 2002;23:13–8.
 131. Dunn A, Rush G. Electrical stimulation in treatment of delayed union and nonunion of fractures and osteotomies. *South Med J*. 1984;77:1530–4.
 132. Foley K, Mroz T, Arnold P. Randomized, prospective, and controlled clinical trial of pulsed electromagnetic field stimulation for cervical fusion. *Spine J*. 2008;8:436–42.
 133. Fourie JA, Bowerbank P. Stimulation of bone healing in new fractures of the tibial shaft using interferential currents. *Physiother Res Int*. 1997;2:255–68.
 134. Freedman LS. Pulsating electromagnetic fields in the treatment of delayed and non-union of fractures: results from a district general hospital. *Injury*. 1985;16:315–7.
 135. Garland D, Holt P, Harrington JT, Caldwell J, Zizic T, Cholewczynski J. A 3-month, randomized, double-blind, placebo-controlled study to evaluate the safety and efficacy of a highly optimized, capacitively coupled, pulsed electrical stimulator in patients with osteoarthritis of the knee. *Osteoarthr Cartil*. 2007;15(6):630–7.
 136. Goodwin C, Brighton C, Guyer R, Johnson J, Light K, Yuan H. A double blind study of capacitively coupled electrical stimulation as an adjunct to lumbar spinal fusion. *Spine*. 1999;24:1349–57.
 137. Hanft JR, Goggin JP, Landsman A, Surprenant M. The role of combined magnetic field bone growth stimulation as an adjunct in the treatment of neuroarthropathy/Charcot joint: an expanded pilot study. *J Foot Ankle Surg*. 1998;37:510–5.
 138. Hannemann P, Göttgens KW, van Wely BJ, Kolkman KA, Werre AJ, Poeze M, et al. Pulsed electromagnetic fields in the treatment of fresh scaphoid fractures. A multicenter, prospective, double blind, placebo controlled, randomized trial. *BMC Musculoskelet Disord*. 2011;12:90. <https://doi.org/10.1186/1471-2474-12-90>.
 139. Hannemann PFW, Göttgens KWA, van Wely BJ, Kolkman KA, Werre AJ, Poeze M, et al. The clinical and radiological outcome of pulsed electromagnetic field treatment for acute scaphoid fractures: a randomised double-blind placebo-controlled multicentre trial. *J Bone Jt Surg Br*. 2012;94(10):1403–8.
 140. Holmes GB. Treatment of delayed unions and nonunions of the proximal fifth metatarsal with pulsed electromagnetic fields. *Foot Ankle Int*. 1994;15:552–6.
 141. Ito H, Shirai Y. The efficacy of ununited tibial fracture treatment using pulsing electromagnetic fields: relation to biological activity on nonunion bone ends. *J Nippon Med Sch*. 2001;68(2):149–53.
 142. Itoh S, Ohta T, Sekino Y, Yukawa Y, Shinomiya K. Treatment of distal radius fractures with a wrist-bridging external fixation: the value of alternating electric current stimulation. *J Hand Surg Eur*. 2008;33(5):605–8. <https://doi.org/10.1177/1753193408092253>.
 143. Jenis L, Howard S, Rebecca C, Brett Y. Prospective comparison of the effect of direct current electrical stimulation and pulsed

- electromagnetic fields on instrumented posterolateral lumbar arthrodesis. *Spinal Disord.* 2000;13:290–6.
144. Jorgensen TE. Electrical stimulation of human fracture healing by means of a slow pulsating, asymmetrical direct current. *Clin Orthop Rel R.* 1977;124:127.
 145. Kahn J. Transcutaneous electrical nerve stimulation for nonunited fractures; a clinical report. *Phys Ther.* 1982;62:840–4.
 146. Kane WJ. Direct current electrical bone growth stimulation for spinal fusion. *Spine.* 1988;13:363–5.
 147. Kucharzyk D. A controlled prospective outcome study of implantable electrical stimulation with spinal instrumentation in a high risk spinal fusion population. *Spine.* 1999;24:465–68.
 148. Lazovic M, Kocic M, Dimitrijevic L, Stankovic I, Spalevic M, Ciric T. Pulsed electromagnetic field during cast immobilization in postmenopausal women with Colles' fracture. *Srp Arh Celok Lek.* 2012;140(9–10):619–24.
 149. Linovitz R, Pathria M, Bernhardt M, Green D, Law M, McGuire R, et al. Combined magnetic fields accelerate and increase spine fusion: a double-blind, randomized, placebo controlled study. *Spine.* 2002;27:1383–9.
 150. Livesley PJ, Muggleston A, Whitton J. Electrotherapy and the management of minimally displaced fracture of the neck of the humerus. *Injury.* 1992;23:323–6.
 151. Madronero A, Pitillas I, Manso FJ. Pulsed electromagnetic field treatment failure in radius non-united fracture healing. *J Biomed Eng.* 1988;10:463–6.
 152. Mammi GI, Rocchi R, Cadossi R, et al. The electrical stimulation of tibial osteotomies: A double-blind study. *Clin Orthop.* 1993;288:246–53.
 153. Marks RA. Spine fusion for discogenic low back pain: outcomes in patients treated with or without pulsed electromagnetic field stimulation. *Adv Ther.* 2000;17:57–67.
 154. Martinez-Rondanelli A, Martinez JP, Moncada ME, Manzi E, Pinedo CR, Cadavid H. Electromagnetic stimulation as coadjuvant in the healing of diaphyseal femoral fractures: a randomized controlled trial. *Colomb Med (Cali).* 2014;45(2):67–71.
 155. Massari L, Fini M, Cadossi R. Biophysical stimulation with pulsed electromagnetic fields in osteonecrosis of the femoral head. *J Bone Jt Surg Am.* 2006;88:56–60.
 156. Masureik C, Eriksson C. Preliminary clinical evaluation of the effect of small electrical currents on the healing of jaw fractures. *Clin Orthop Relat R.* 1977;124:84–91.
 157. Meskens M, Stuyck J, Mulier J. Treatment of delayed union and nonunion of the tibia by pulsed electromagnetic fields. *Bull Hosp Jt Dis Orthop Inst.* 1988;48:170–5.
 158. Paterson D, Simonis RB. Electrical stimulation in the treatment of congenital pseudoarthrosis of the tibia. *J Bone Jt Surg Br.* 1985;67:454–62.
 159. Punt BJ, Den Hoed PT, Fontijne WPJ. Pulsed electromagnetic fields in the treatment of nonunion. *Eur J Orthop Surg Traumatol.* 2008;18:127–33.
 160. Reilingh ML, van Bergen CJA, Gerards RM, van Eekeren IC, de Haan RJ, Sierevelt IN, et al. Effects of pulsed electromagnetic fields on return to sports after arthroscopic debridement and microfracture of osteochondral talar defects: a randomized, double-blind, placebo-controlled, multicenter trial. *Am J Sports Med.* 2016;44(5):1292–300. <https://doi.org/10.1177/0363546515626544>.
 161. Rogozinski A, Rogozinski C. Efficacy of implanted bone growth stimulation in instrumented lumbosacral spinal fusion. *Spine.* 1996;21:2479–483.
 162. Saltzman C, Lightfoot A, Amendola A. PEMF as treatment for delayed healing of foot and ankle arthrodesis. *Foot Ankle Int.* 2004;25:771–3.
 163. Scott G, King JB. A prospective, double-blind trial of electrical capacitive coupling in the treatment of non-union of long bones. *J Bone Jt Surg Am.* 1994;76:820–6.
 164. Shi H, Xiong J, Chen Y, Wang J, Qiu X, Wang Y, et al. Early application of pulsed electromagnetic field in the treatment of postoperative delayed union of long-bone fractures: a prospective randomized controlled study. *BMC Musculoskelet Disord.* 2013;14:35. <https://doi.org/10.1186/1471-2474-14-35>.
 165. Simmons JW. Treatment of failed posterior lumbar interbody fusion (PLIF) of the spine with pulsing electromagnetic fields. *Clin Orthop Relat Res.* 1985;183:127.
 166. Simmons JW, Mooney V, Thacker I. Pseudarthrosis after lumbar spine fusion: nonoperative salvage with pulsed electromagnetic fields. *Am J Orthop.* 2004;33:27–30.
 167. Streit A, Watson BC, Granata JD, Philbin TM, Lin H-N, O'Connor JP, et al. Effect on clinical outcome and growth factor synthesis with adjunctive use of pulsed electromagnetic fields for fifth metatarsal nonunion fracture: a double-blind randomized study. *Foot Ankle Int.* 2016;37(9):919–23. <https://doi.org/10.1177/1071100716652621>.
 168. Steinberg ME, Brighton CT, Bands RE, Hartman KM. Capacitive coupling as an adjunctive treatment for avascular necrosis. *Clin Orthop Relat Res.* 1990;261:11–8.
 169. van Bergen CJA, Blankevoort L, de Haan RJ, Sierevelt IN, Meuffels DE, d'Hooghe PRN, et al. Pulsed electromagnetic fields after arthroscopic treatment for osteochondral defects of the talus: double-blind randomized controlled multicenter trial. *BMC Musculoskelet Disord.* 2009;10:83. <https://doi.org/10.1186/1471-2474-10-83>.
 170. Wahlstrom O, Knutsson H. A device for generation of electromagnetic fields of extremely low frequency. *J Biomed Eng.* 1984;6:293–6.
 171. Welch WC, Willis SL, Gerszten PC. Implantable direct current stimulation in para-axial cervical arthrodesis. *Adv Ther.* 2004;21:389–400.